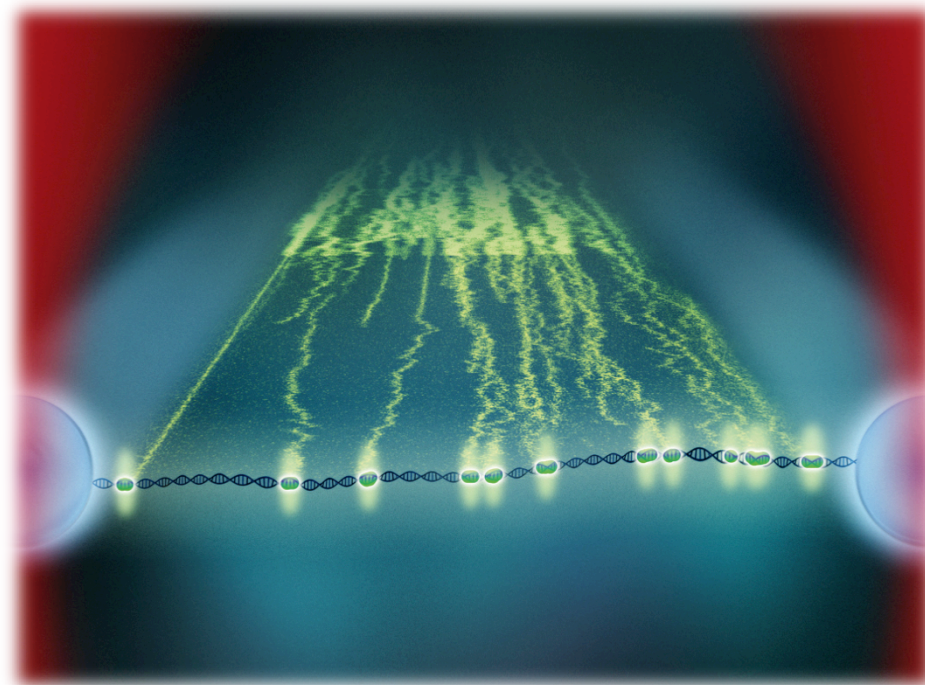


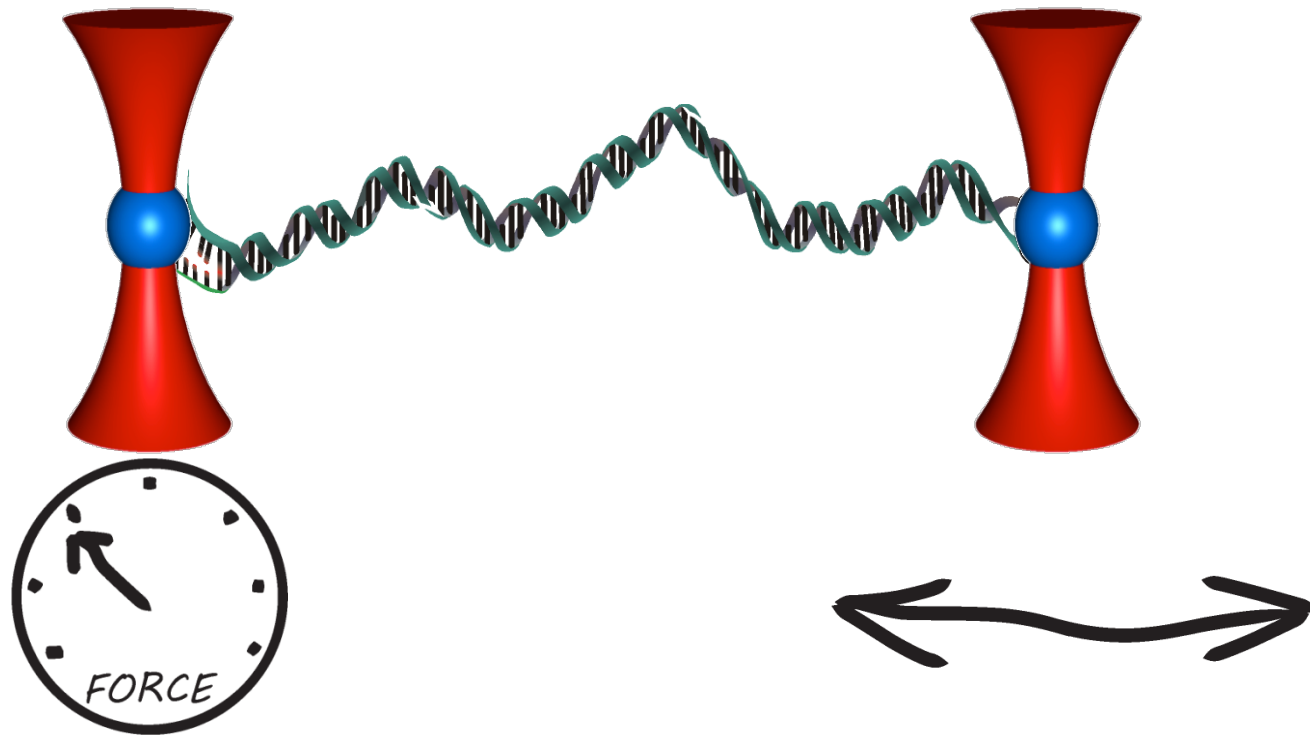
Correlative Tweezers Fluorescence Microscopy: super-resolution STED-imaging of DNA-protein interactions

Erwin J.G. Peterman

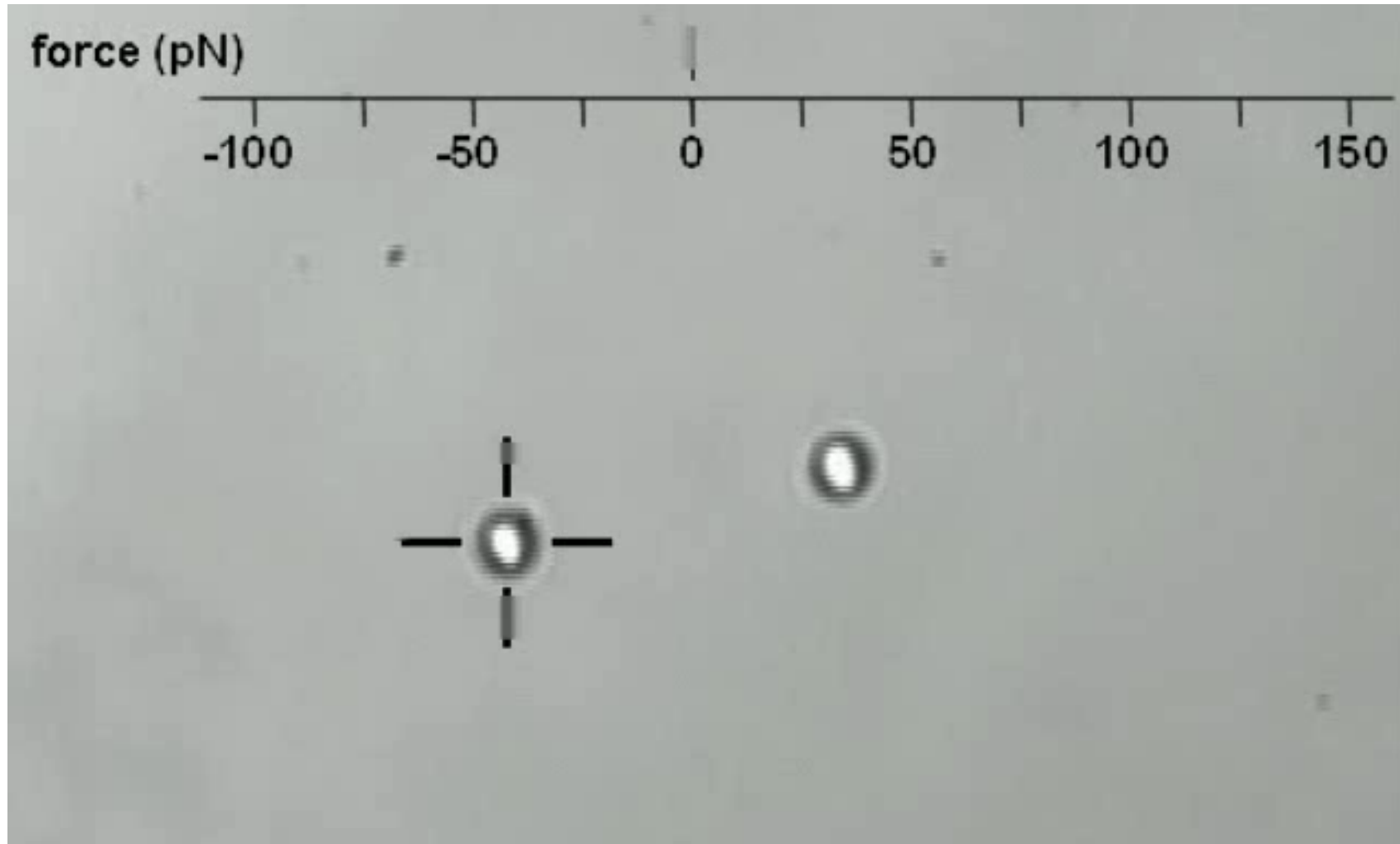
VU University Amsterdam, The Netherlands



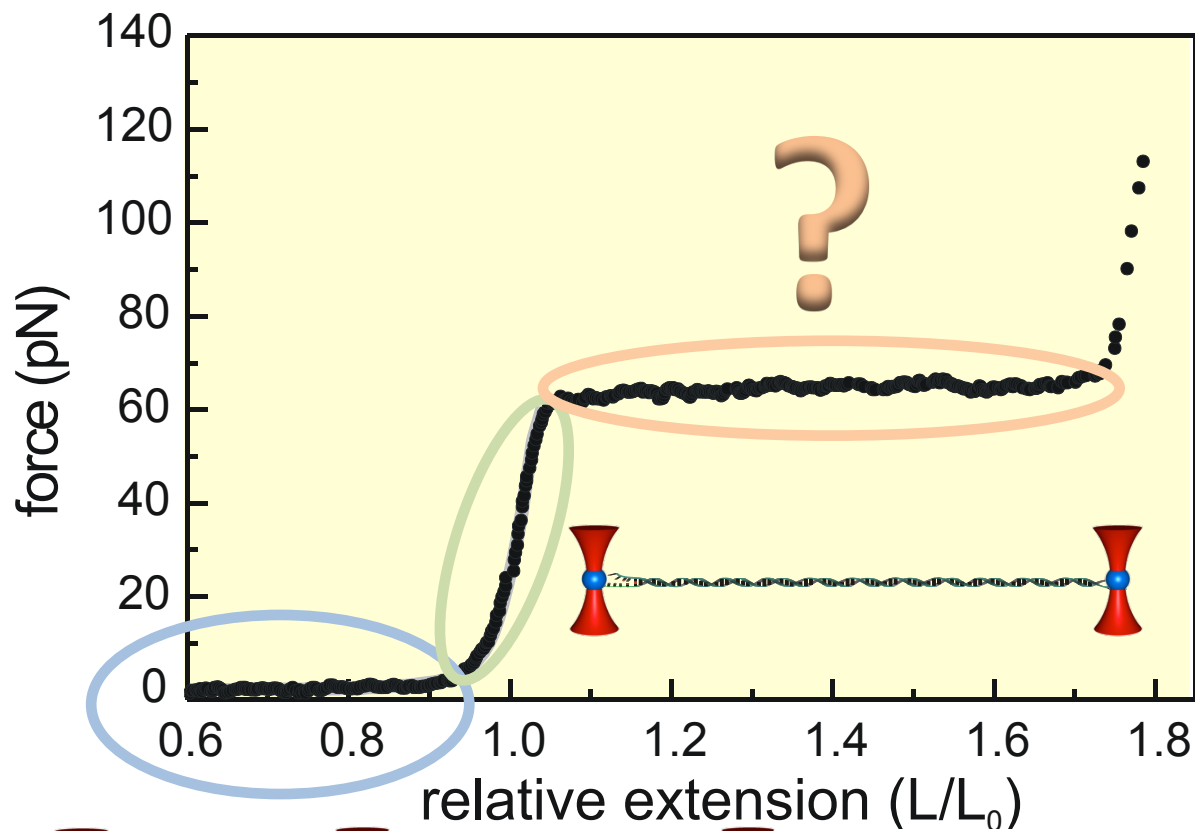
Manipulation of DNA with optical tweezers



Manipulation of DNA with optical tweezers



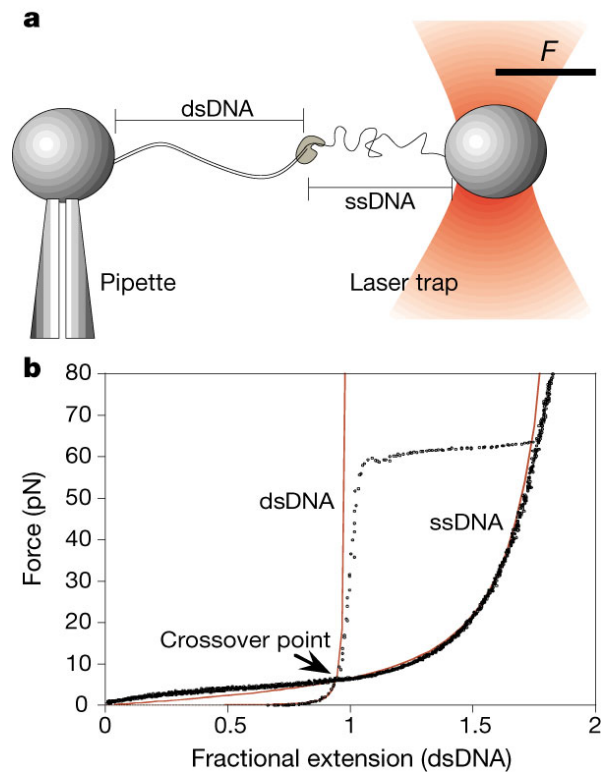
Manipulation of DNA with optical tweezers



global information on whole DNA molecule

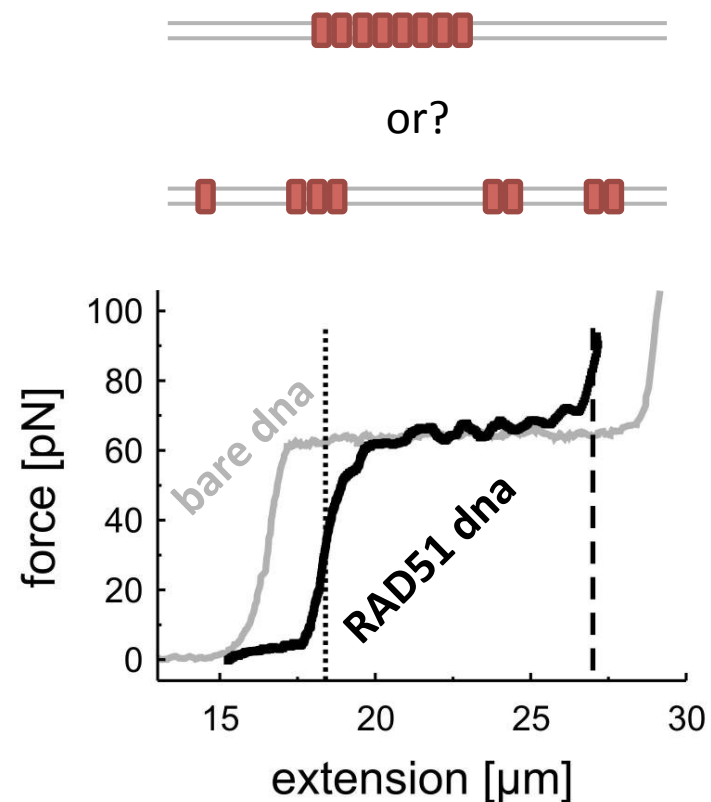
Manipulation of DNA with optical tweezers

Applications



DNAp converting ssDNA in dsDNA

Wuite et al., Nature 2000

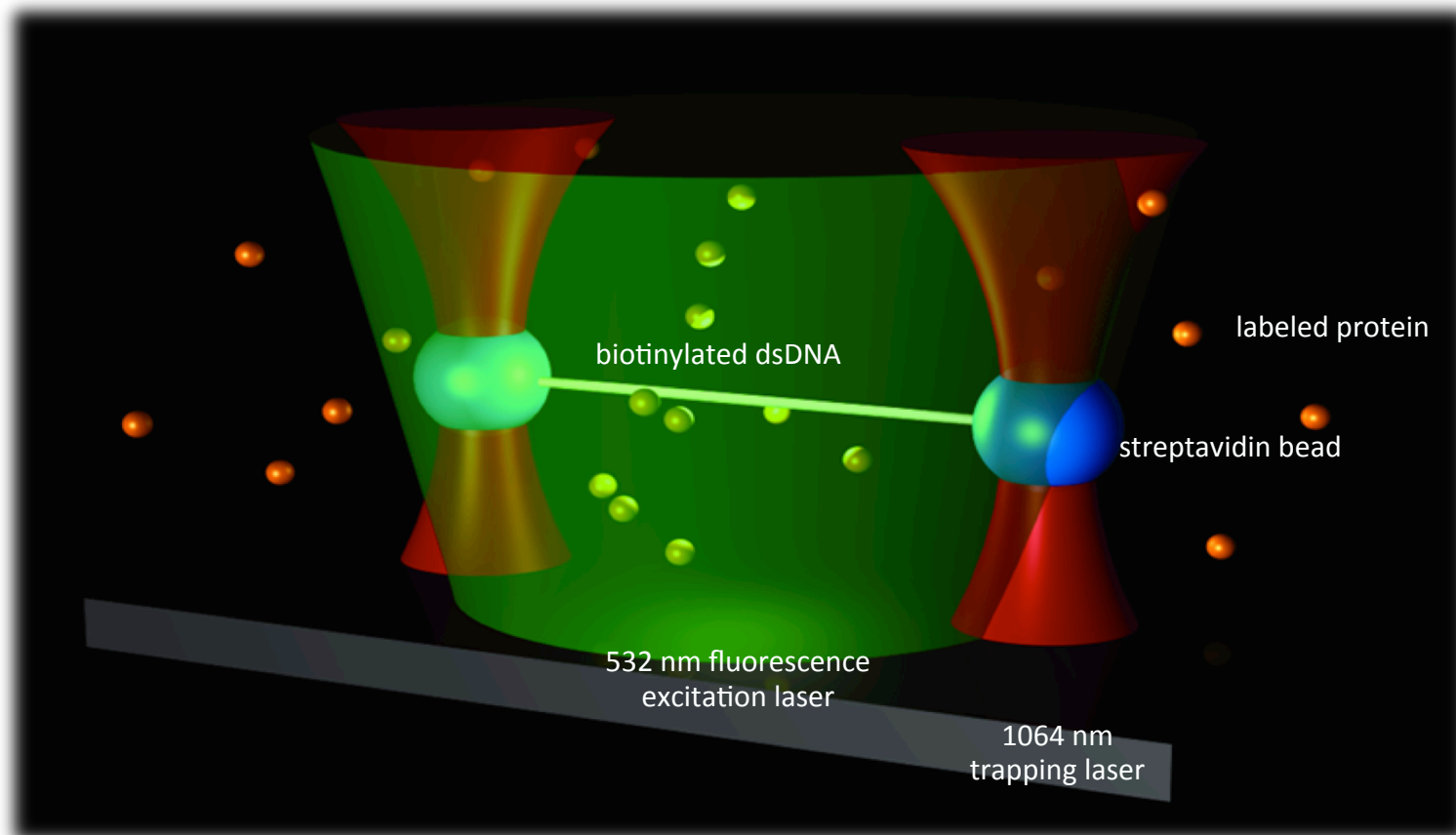


RAD51 binding to dsDNA

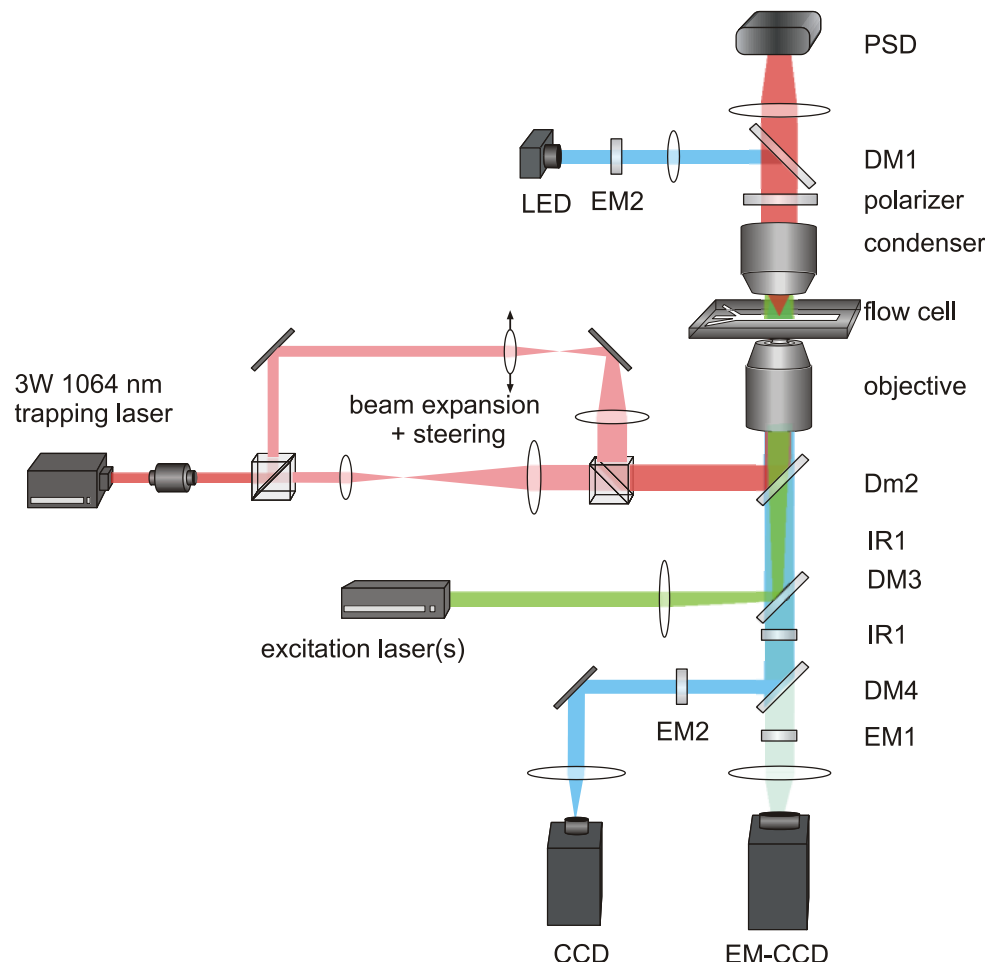
van Mameren et al., Biophys J 2006

Heller et al., Chemical Reviews 2014

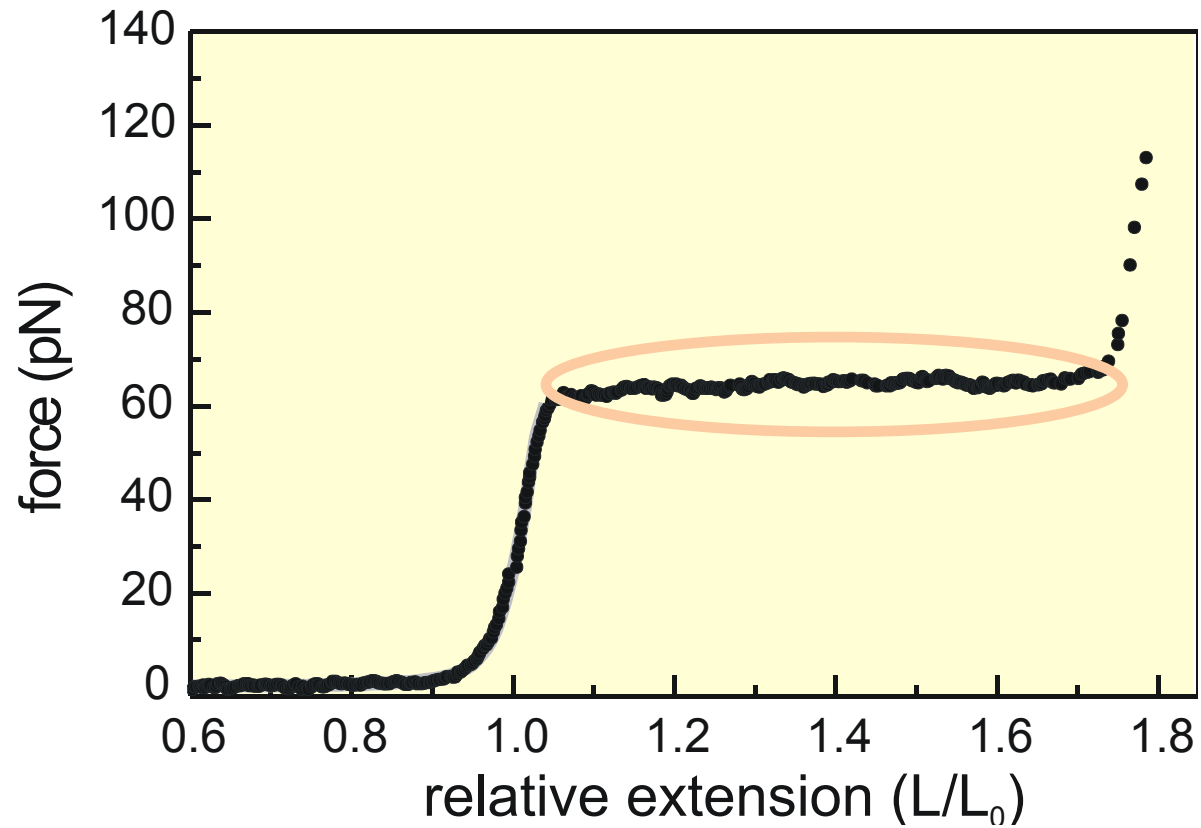
Our approach: combine optical tweezers with fluorescence microscopy



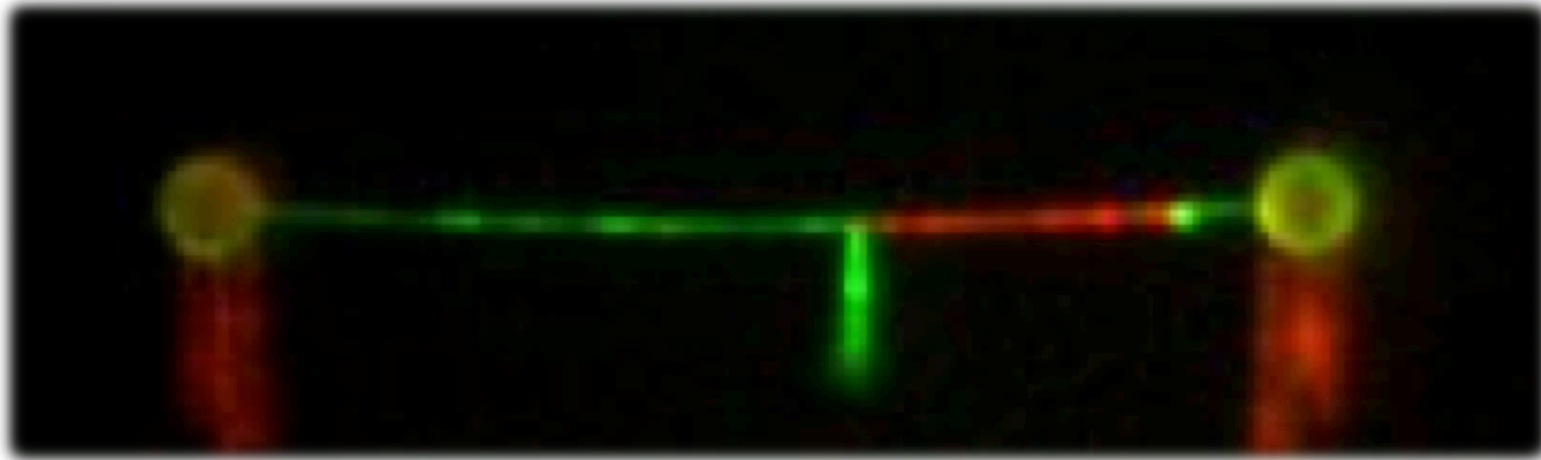
Optical tweezers combined with epi wide-field fluorescence microscopy



Overstretching = force-induced melting*

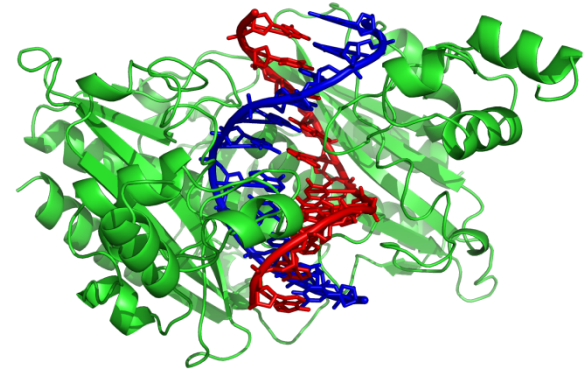


Overstretching = force-induced melting*



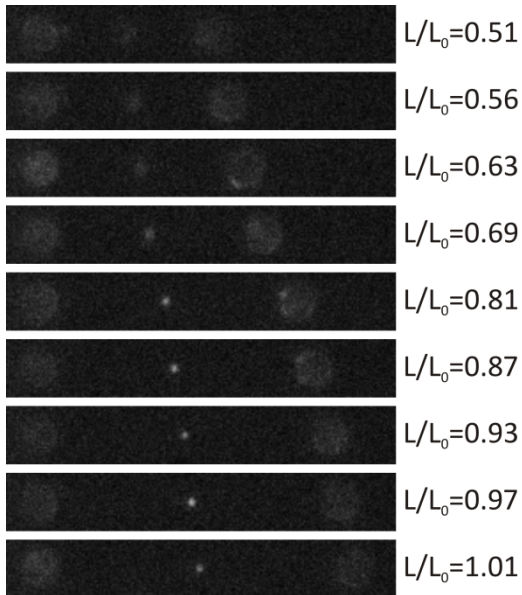
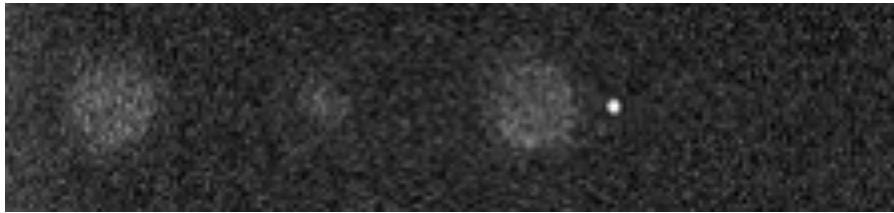
- **green** eGFP-RPA ssDNA
- **red** POPO-3 dsDNA
- buffer flow stretches frayed ssDNA

Single-molecule sensitivity in fluorescence

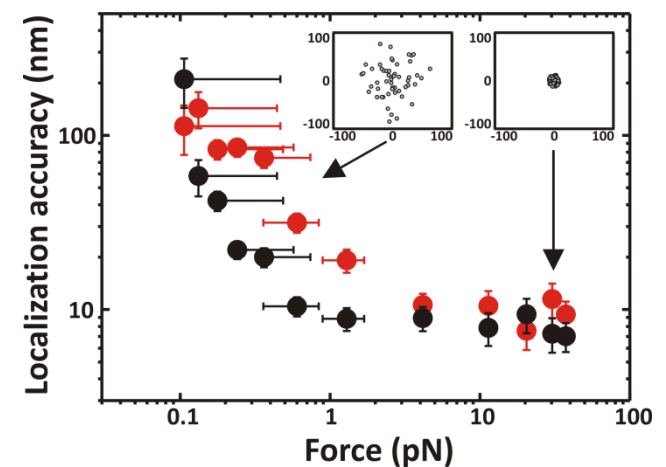
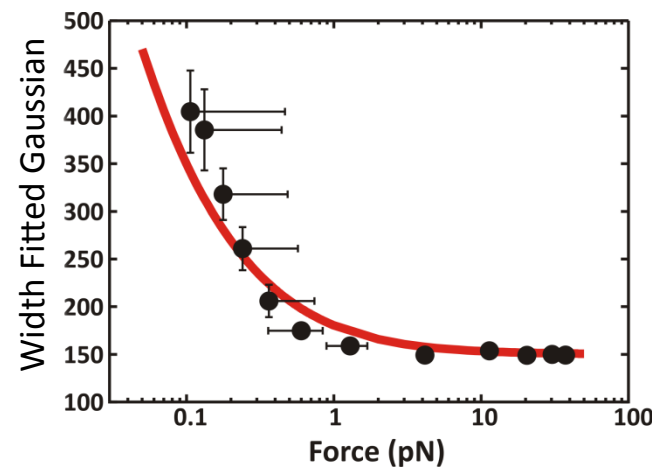
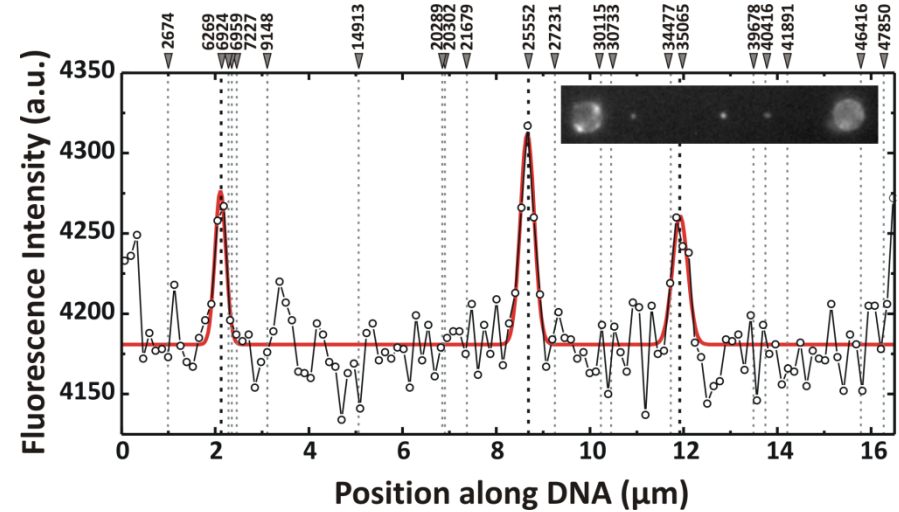


- Alexa555-labeled EcoRV
- Single-step bleaching → single molecules

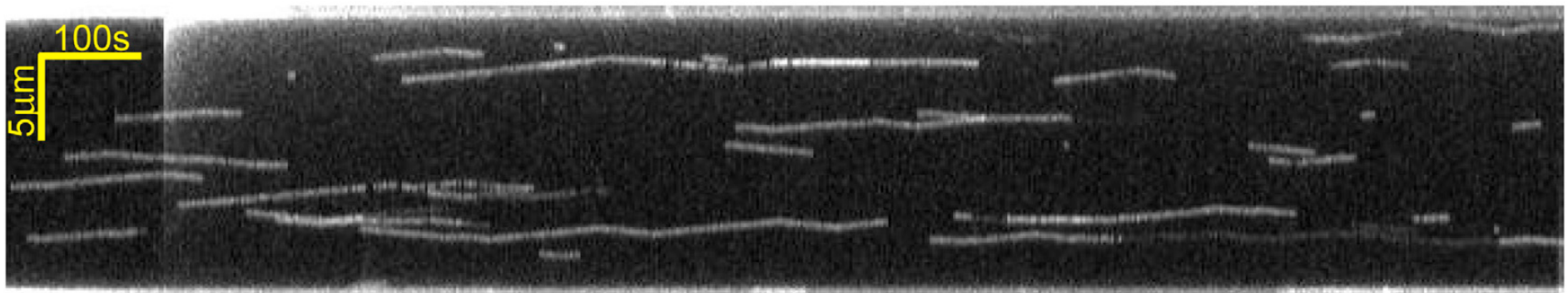
Single-molecule sensitivity in fluorescence



EcoRV binding: error with known sites 14 ± 7 nm



Example: PICH, a DNA translocase specially adapted for processing anaphase bridge DNA



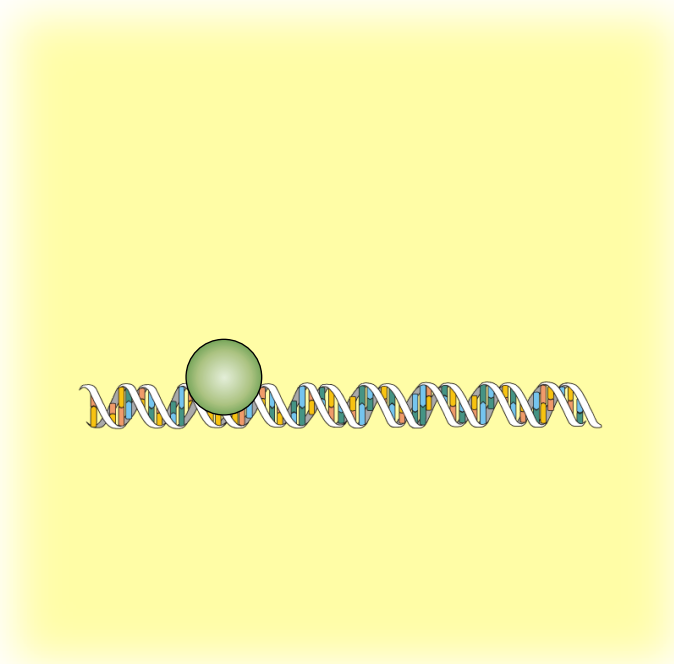
$$v = 10 \text{ nm/s} \approx 30 \text{ bp/s}$$

Limitations

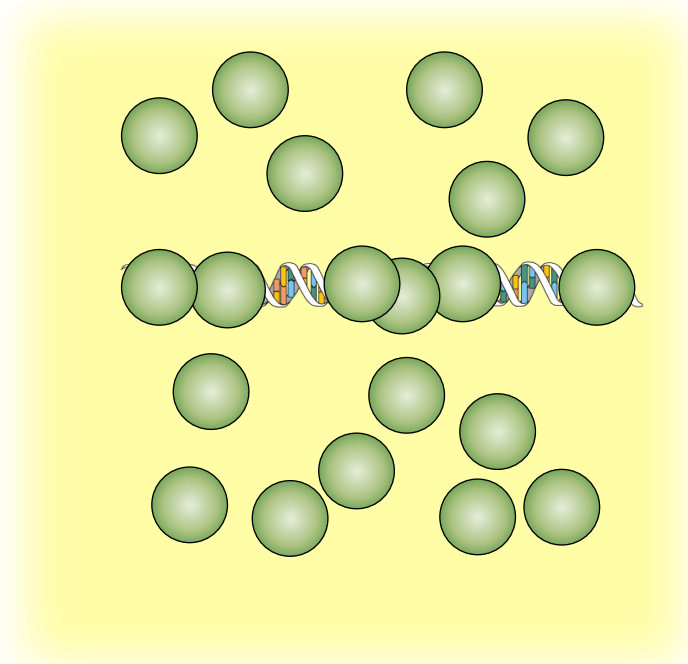
in vitro

vs

in vivo

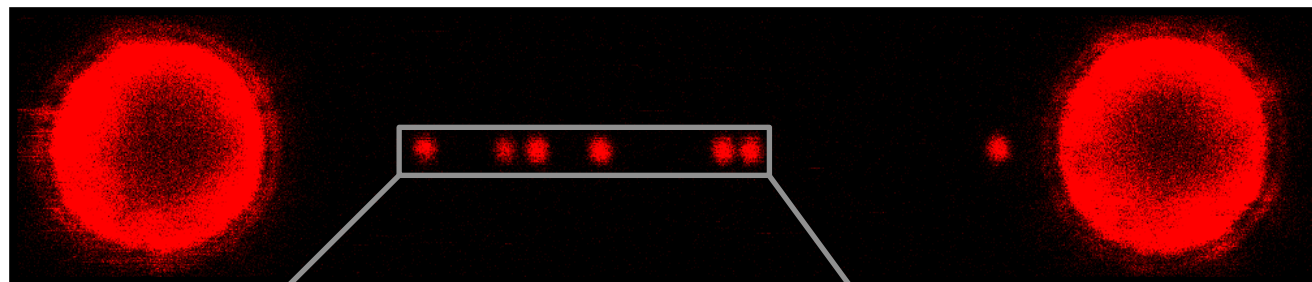
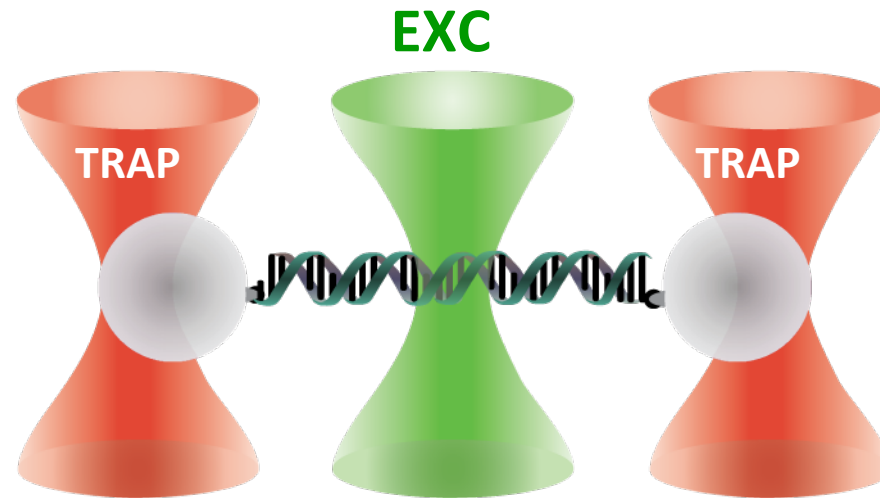


1. Low conc. to suppress background
2. Low conc. to observe separate binding



1. High concentration
2. High density on DNA

Confocal fluorescence microscopy



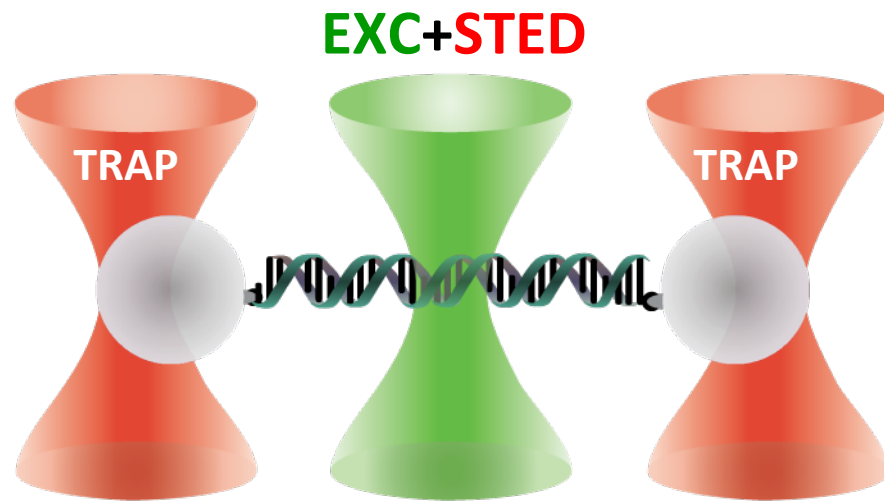
Atto647N-protein



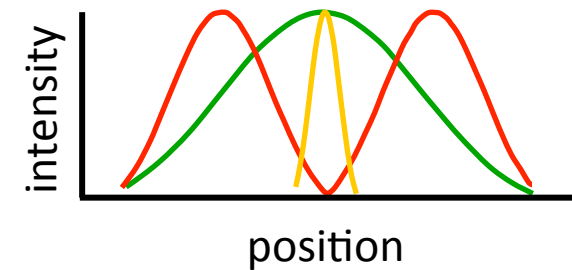
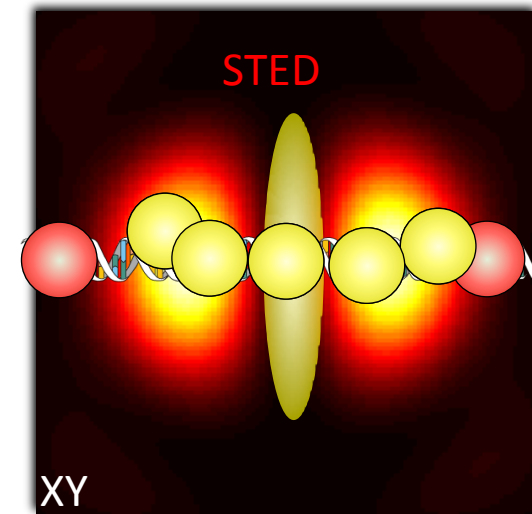
+ 40 nM Atto647N

Limit: 100 nM (at 1 ms)

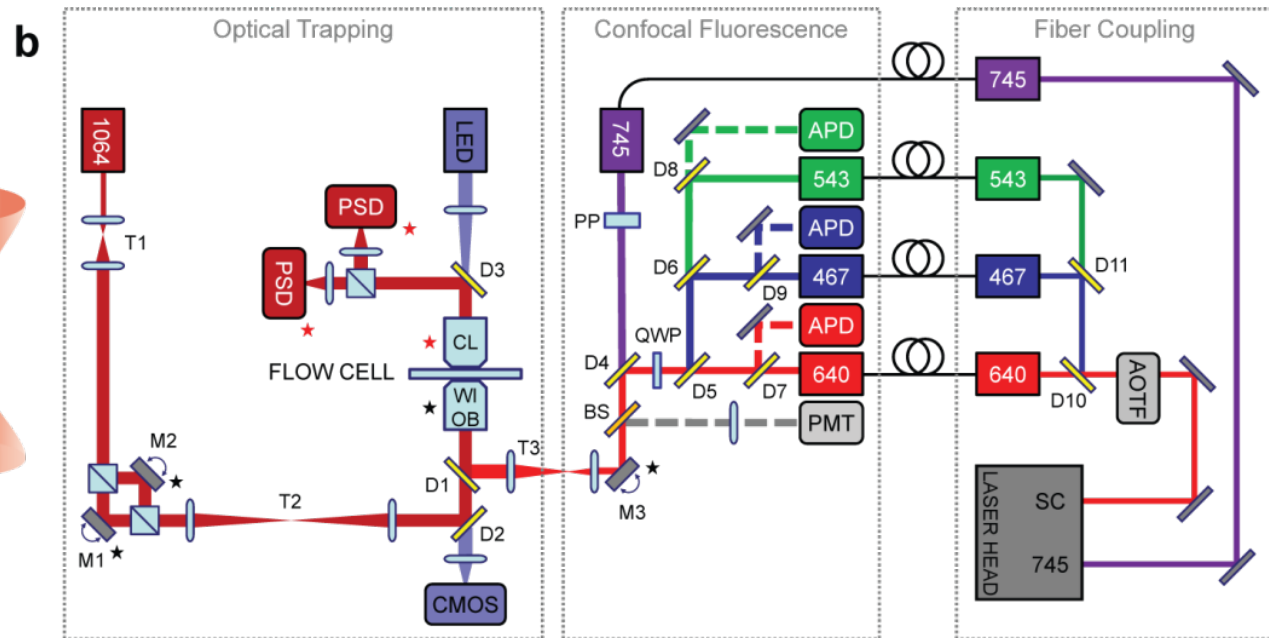
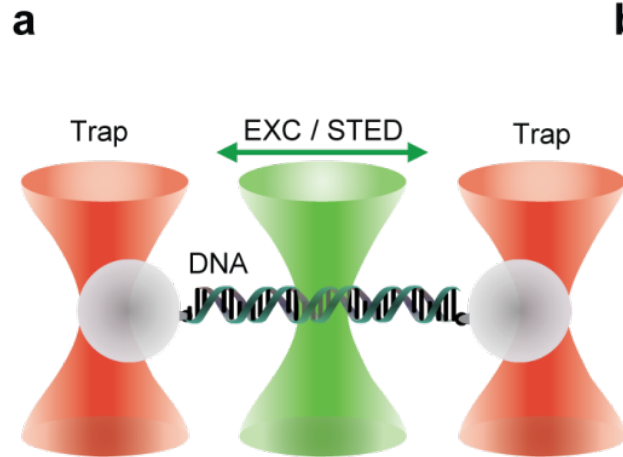
1D stimulated-emission depletion



High density **ON** DNA

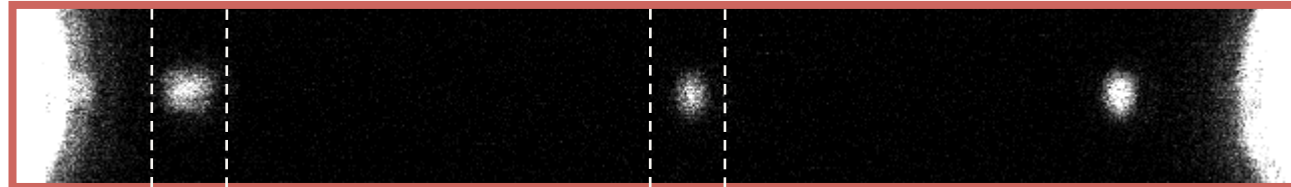


1D stimulated-emission depletion

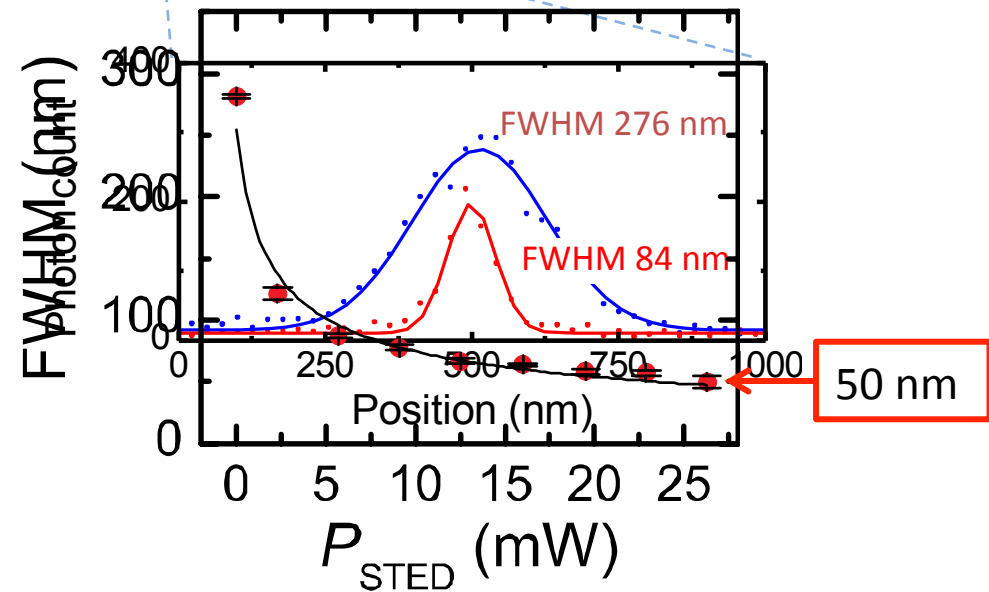
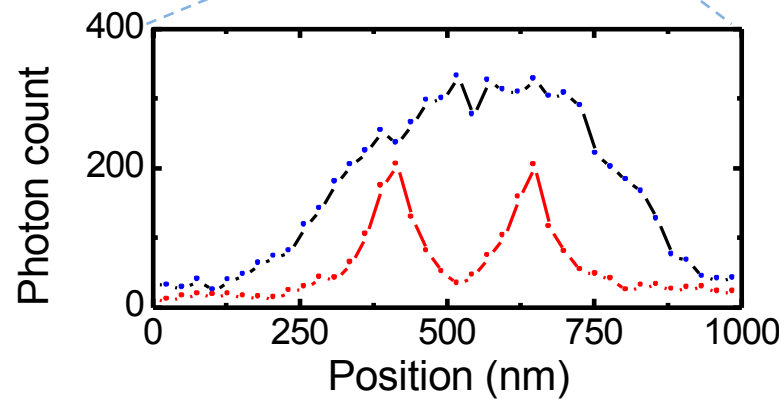
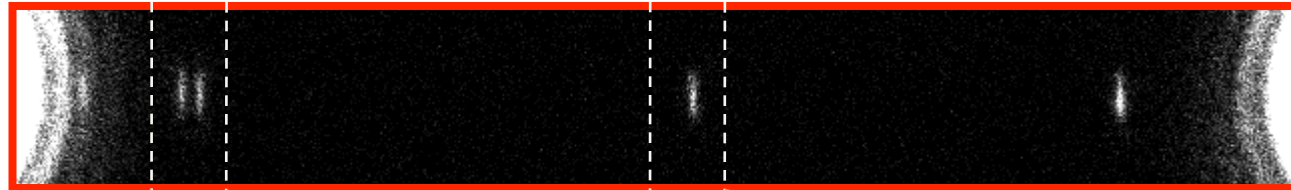


1D stimulated-emission depletion

Confocal



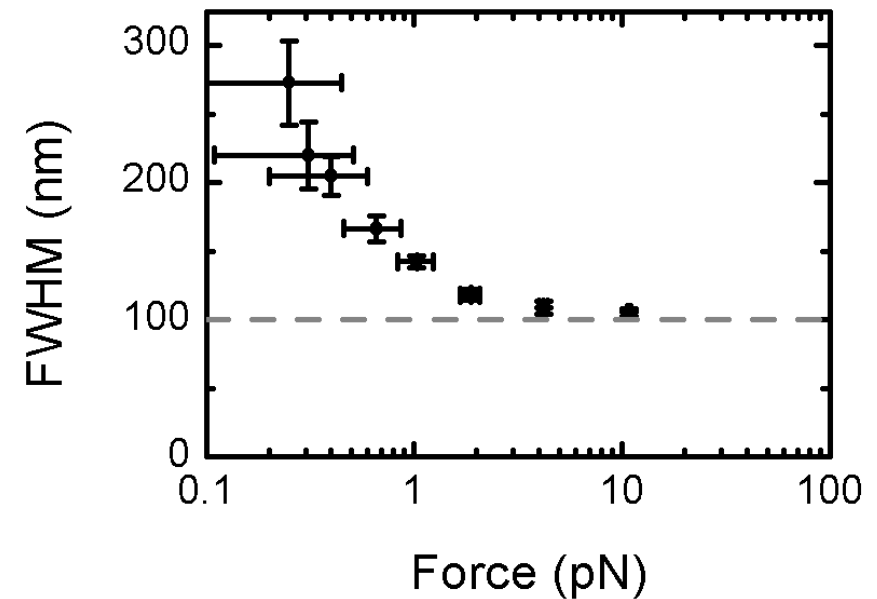
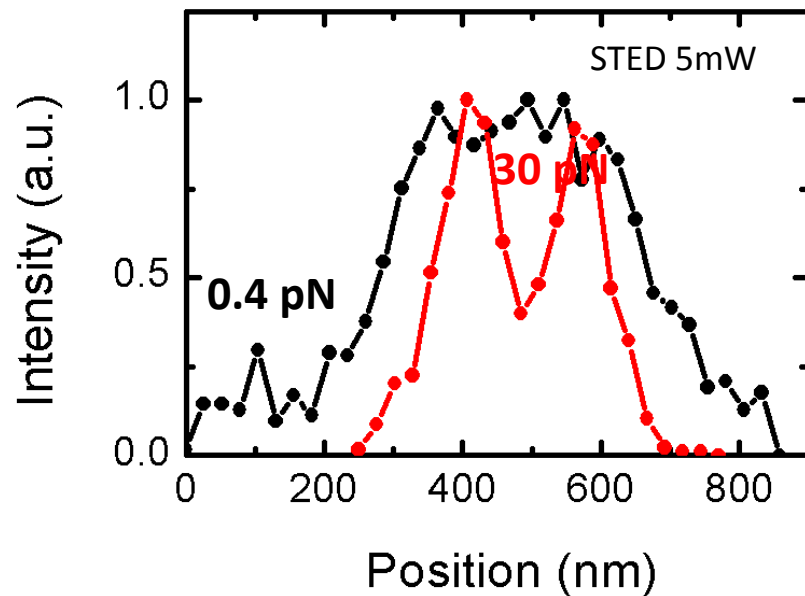
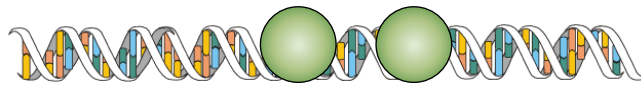
STED



Exc: 640nm 4uW, STED: 745nm 5mW [20MHz, 100ps]

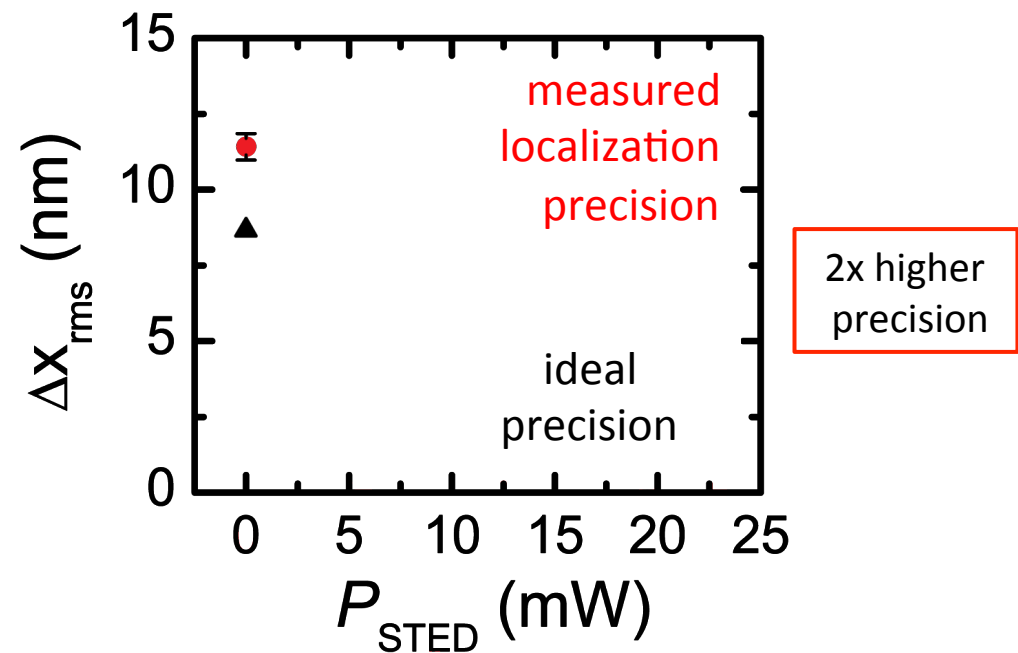
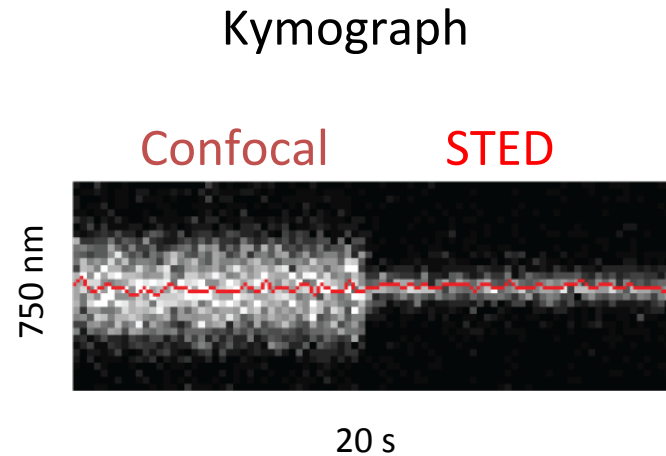
1D stimulated-emission depletion

Point-spread function tension dependent

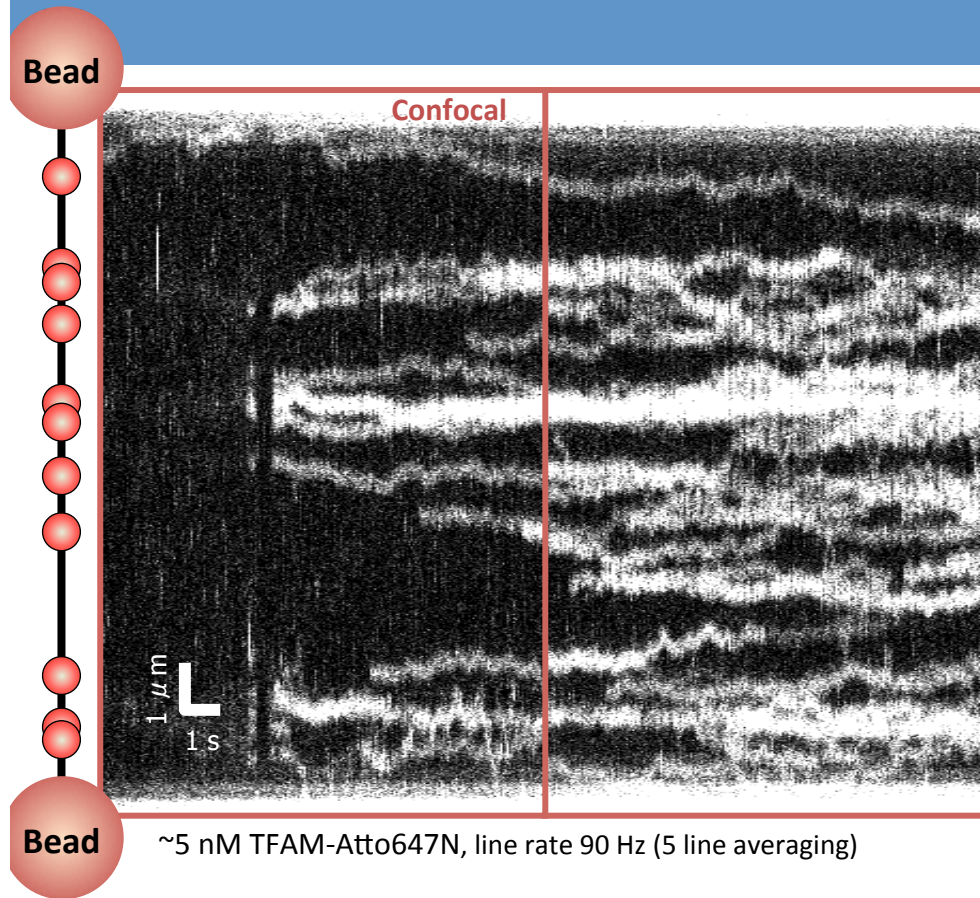


1D stimulated-emission depletion

Localization accuracy

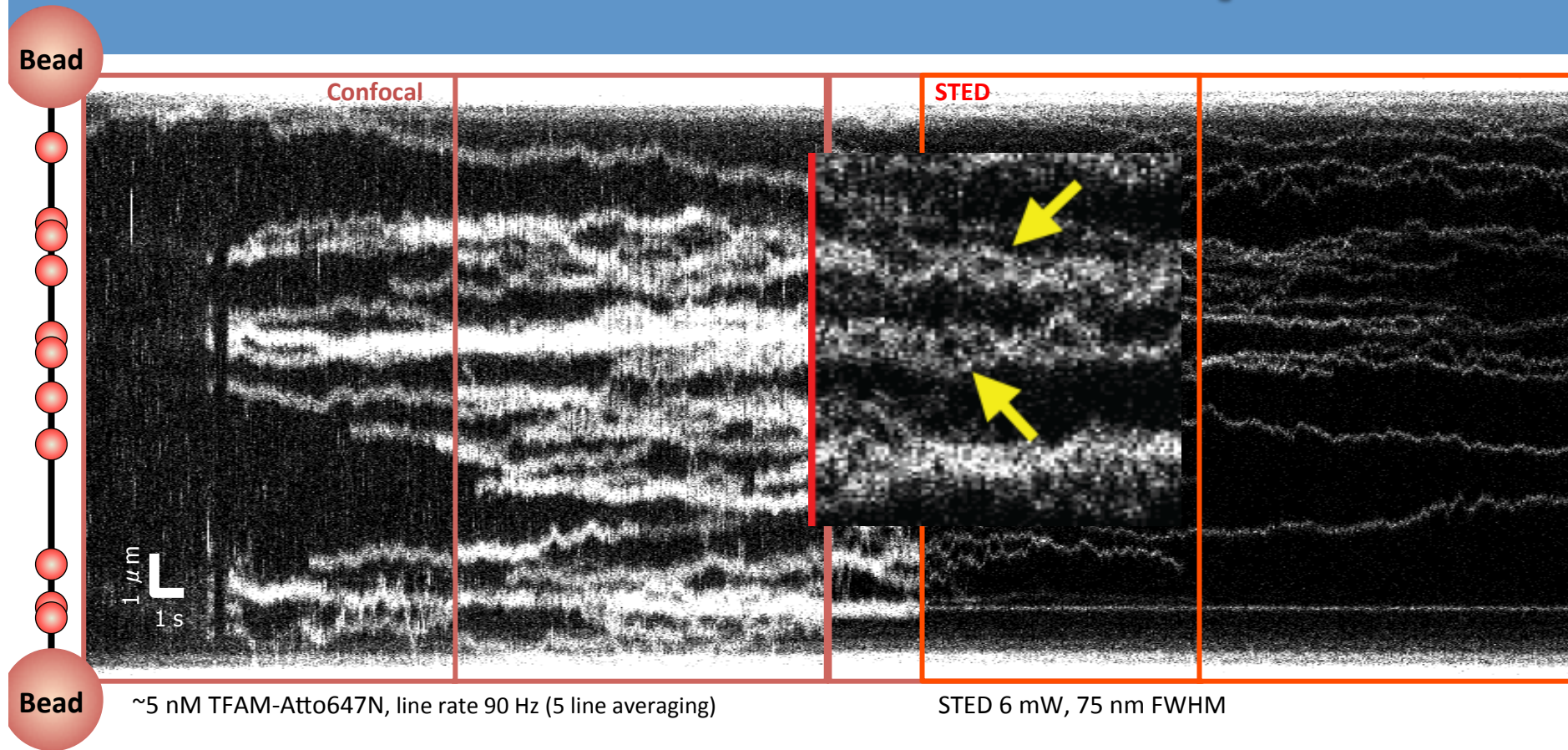


1D stimulated-emission depletion



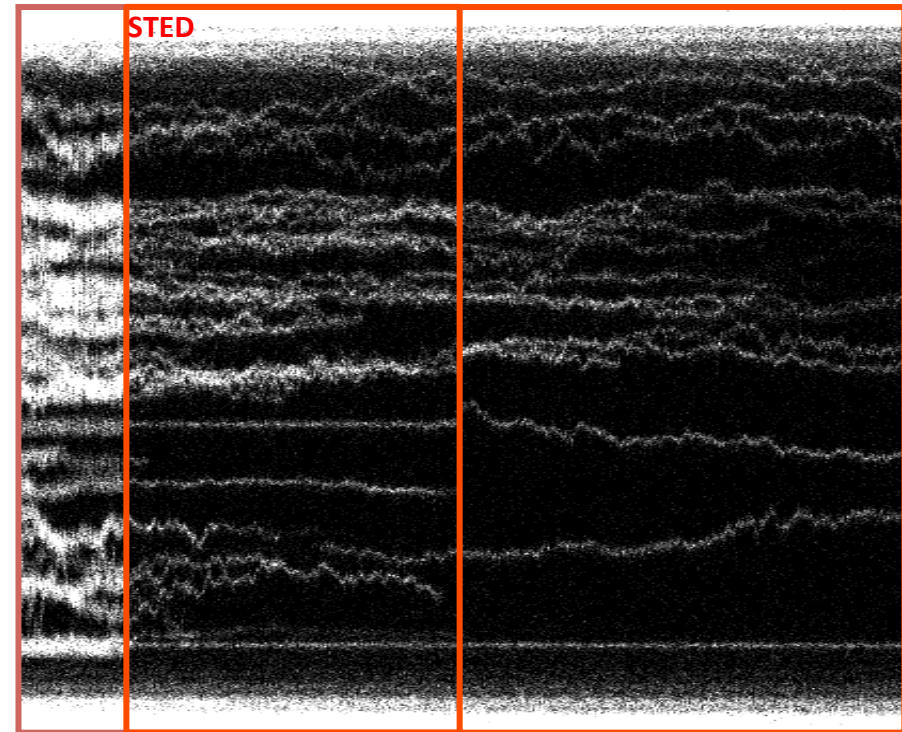
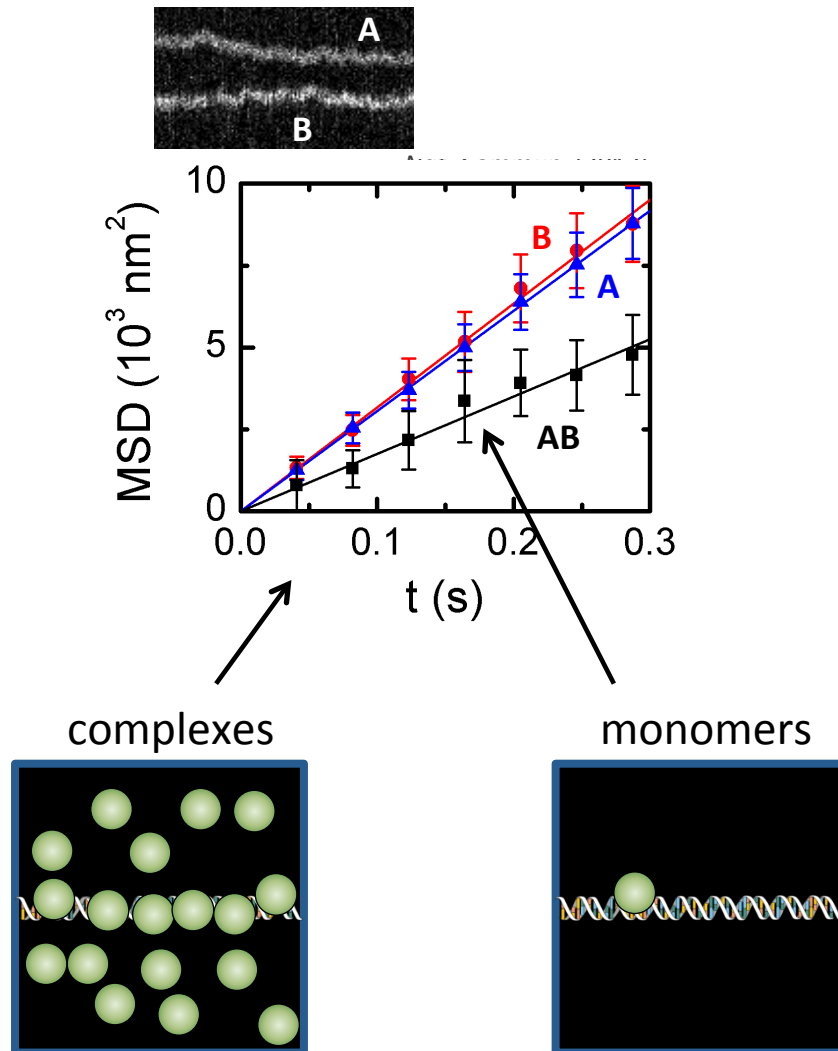
Application to TFAM (Mitochondrial Transcription Factor A)

1D stimulated-emission depletion



Application to TFAM (Mitochondrial Transcription Factor A)

1D stimulated-emission depletion



STED 6 mW, 75 nm FWHM

Conclusions

- Fluorescence microscopy adds local information to tweezers experiments.
- Super-resolution localization accuracy can be achieved using epi wide-field excitation.
- Better suppression of background due to fluorophores in solution using confocal.
- Super resolution (so far down to ~ 50 nm) using STED, localization accuracy ~ 7 nm.

LUMICKS

Correlative optical tweezers-fluorescence microscopy systems

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Acknowledgements

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